Effect of Crizanlizumab on Blood Cell Adhesion in Sickle Cell Disease: A Standardized **Microfluidic Assessment via the Endothelium-on-a-Chip**



INTRODUCTION

- > Chronic upregulation of P-selectin on blood cells and the endothelium leads to abnormal red blood cell (RBC) adhesion to endothelial cells, significantly contributing to vaso-occlusive crises (VOCs), which are a major cause of morbidity and mortality in patients with sickle cell disease (SCD). Crizanlizumab (a.k.a. SEG101) is a humanized anti-P-selectin monoclonal antibody and has recently been approved by the Food and Drug Administration to reduce the frequency of VOCs in SCD patients.
- > There is currently no existing feasible clinical in vitro model for monitoring the patient-specific effects of Crizanlizumab on cellular adhesion, which we would argue is largely due to the lack of a universally accepted, standardized physiologic flow-based adhesion assay with which to measure blood cell adhesion to human endothelial cells. Utilizing such an *in vitro* assay would greatly help visualize cellular adhesion before and after therapeutic interventions and may reveal patient-specific responses to combination therapies.
- \succ To this end, we have developed a standardized endothelialized microfluidic platform: **Endothelium-on-a-chip**, for in vitro assessment of the effect of Crizanlizumab on patient-specific RBC adhesion to heme-activated human endothelial cells. We have previously shown that RBC adhesion to heme activated endothelial cells may correlate with a more severe disease phenotype.

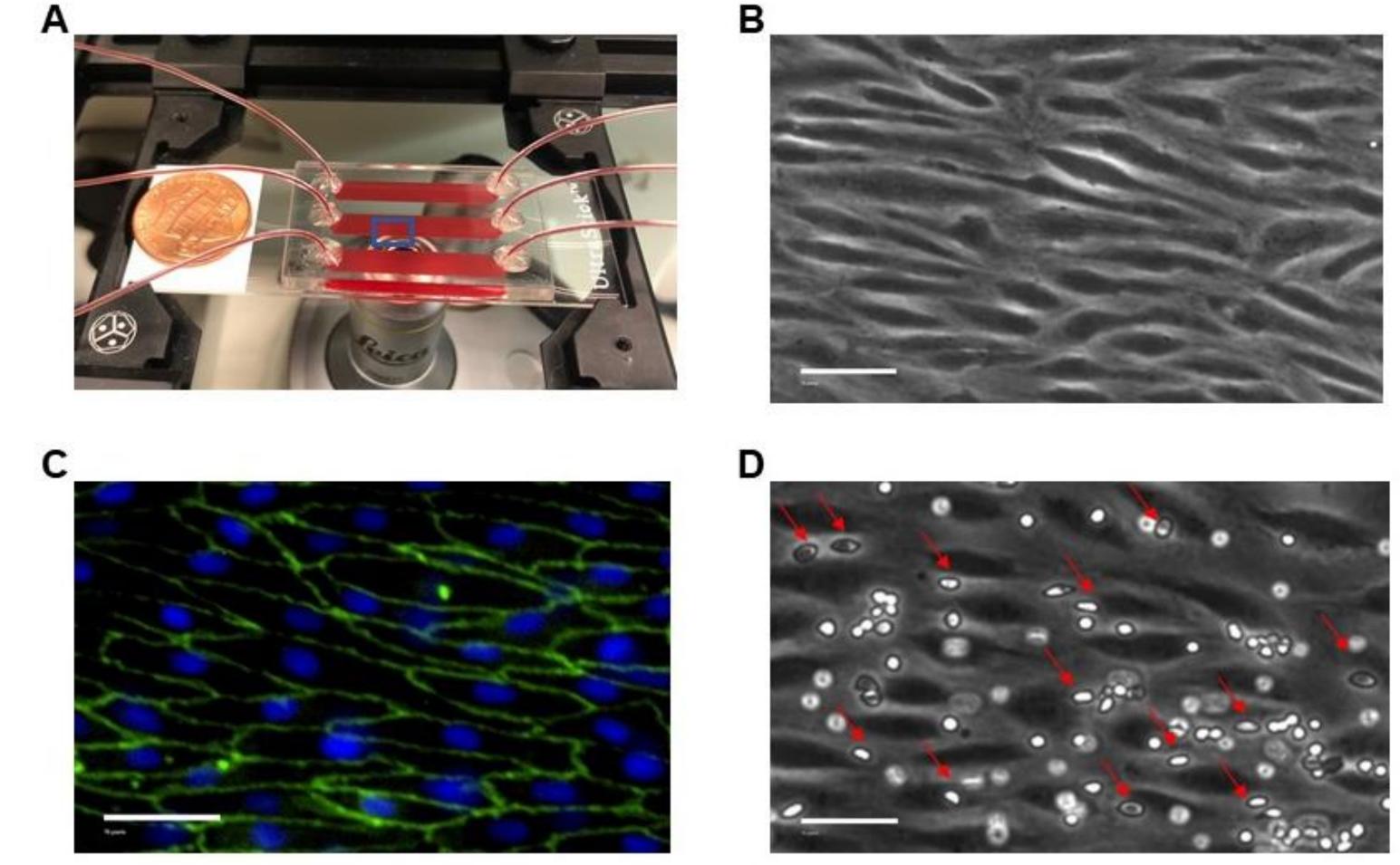


Figure 1. The proposed microfluidic system functionalized with HUVECs. (A) Endothelium-on-a-chip set-up. The phase-contrast (B) and fluorescent (C) images of HUVEC demonstrate a confluent layer of cells aligned with flow (Hoechst blue, CD144 green). (D) RBCs adhesion to Heme activated HUVECs. Microscope images at 10X. Scale bare 50um

MATERIALS & METHODS

- > Microfluidic channels were fabricated via a lamination technique using laser micro-machined components. The device assembly was performed by placing a laser-cut double sided adhesive (DSA) film in between an acrylic top cover, that includes the inlet and outlet ports, and a glass microscope slide (Fig.1A). The height of the microchannels was determined by DSA thickness, which was 50 µm mimicking the size scale of human postcapillary venules.
- > To prepare endothelialized microchannels, Human Umbilical Vein Endothelial Cells (HUVECs) were cultured and seeded into fibronectin-functionalized microchannels at a density of 8x10⁶ cells/mL and incubated in static conditions for 2 hours at 37°C and 5% CO₂.
- \succ The endothelialized microchannels were then cultured under flow (~15 dyne/cm²) for 48-hours at 37°C and 5% CO_2 prior to adhesion experiments. Figure 1 B and C show the HUVEC aligned in the channel.
- > Adhesion experiments were performed for two different endothelial activation durations: 4 hours (long-term) and 30 minutes (short-term). For long-term activation, HUVECs were treated with 40 μ M of heme for 4 hours +/- 100 µg/ml Crizanlizumab for 1 hour followed by injection of blood samples through the microfluidic channels. For short-term activation, blood samples were supplemented with 40 μ M heme +/- 100 μ g/ml Crizanlizumab and injected through the microfluidic channels for 15 minutes. Thereafter, non-adherent RBCs were rinsed via either only 40 μM heme-containing basal medium or 40 μM heme- and 100 μg/ml Crizanlizumab-containing basal medium for another 15 minutes.
- > Whole blood samples were collected from 19 subjects with SCD (18 HbSS and 1 HbSC) in EDTA vacutainers. RBCs were isolated via centrifugation from whole blood and then resuspended in basal cell culture medium at a hematocrit of 20% buffered with 10 mM of HEPES.

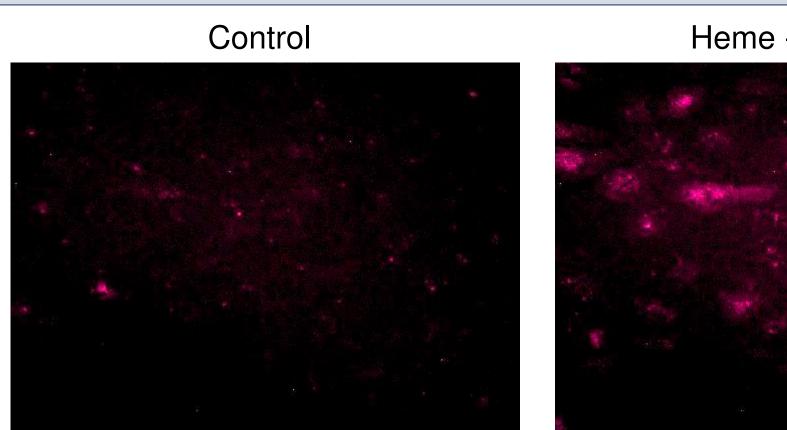
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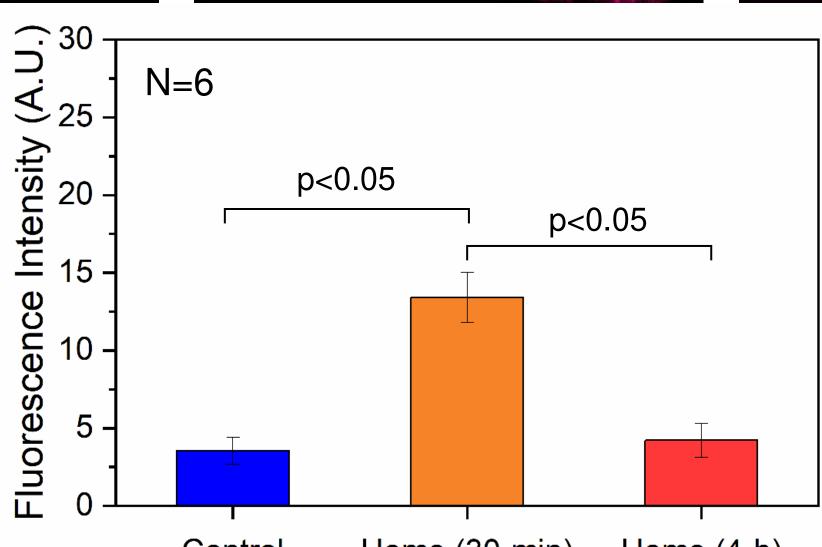
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- injecting the samples.
- dyne/cm². We then quantified the number of adherent RBCs at min=0 and min=15.

RESULTS AND DISCUSSION





Control

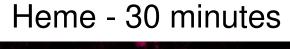
Figure 2. P-selectin surface expression on HUVECs in response to heme activation. Short term heme activation led to a statistically significant increase in surface P-selectin presence while increasing the activation duration to 4 hours lowered the amount of surface P-selectin expression. Fluorescence intensity was quantified using ImageJ.

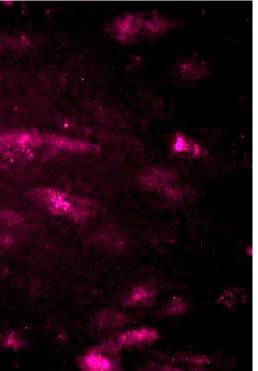
- > Short term (30-min) heme activation significant increase endothelial surface P-selectin expression based on ICAM-1 expression levels following a 4-hours heme activation (data not shown).
- \succ 4-hours heme activation of HUVECs resulted in statistically significant elevated RBC adhesion compared to p<0.05).
- \succ Treatment of 4-hours heme-activated HUVECs with Crizanlizumab did not statistically significant decrease Crizanlizumab treated HUVECs for certain subjects (**Fig. 3B**).
- > Crizanlizumab treatment statistically significant reduced the number of adherent RBCs to 15-minute hemebaseline adhesion levels (Fig. 3D).
- channels without the addition of Crizanlizumab.
- of RBCs to the TNF α and heme activated HUVECs.
- therapies in SCD patients during drug development and in clinical trials.

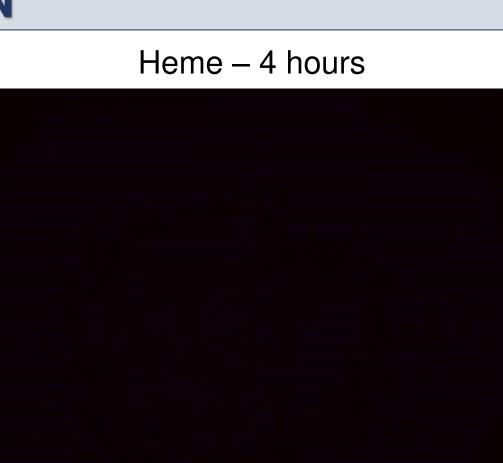
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> For the long-term activation experiments, isolated RBCs were directly injected over endothelial cells. For the short-term activation experiments, heme and Crizanlizumab were added into this RBC suspension prior to

> Detachment experiments were also conducted in this study to determine whether Crizanlizumab would impact already established adhesive events between RBCs and endothelial cells (a.k.a. "crisis-resolving" effect). To do so, RBCs were first allowed to adhere to heme-activated HUVECs (for 15-min) and then the microchannels were rinsed via either a heme- or both heme- and Crizanlizumab-containing solution for another 15 minutes at 1







Heme (30-min) Heme (4-h)

our immunofluorescence studies as shown in Figure 2. Long term (4-hour) heme activation brings the surface P-selectin expression down to the basal level while we observed statistically significant elevated E-selectin and

baseline while adhesion levels were heterogenous among the patient population (Fig. 3A, 1671±522 vs 17±4,

RBC adhesion (Fig. 3A, 1170±413 vs 1671±522, p>0.05), while we observed lower RBC adhesion to

activated HUVECs (Fig. 3C, 135±40 vs 1513±617, p<0.05). Crizanlizumab treatment of HUVECs was able to decrease RBC adhesion for each patient who was tested while certain patients did not have considerable

> As shown in Figure 3E, perfusion of heme-containing wash medium supplemented with Crizanlizumab resulted in higher RBC detachment rates compared to heme-only medium for all three subjects. On average, the percentage of remaining RBCs after a 15-minute wash was only 10% when the heme-containing solution included 100 µg/ml Crizanlizumab. By contrast, more than half of the RBCs still remained in the microfluidic

> In Figure 3F the HUVECs were activated with TNF α (20 ng/ml) for 4 hours previous the short-term (30 minutes) treatment with heme in presence or not of the Crizanlizumab. Crizanlizumab decrease the adhesion

> Our results show that the magnitude of inhibition of RBC adhesion to HUVECs with Crizanlizumab is influenced by the duration of heme-activation (4 hours vs 15 minutes). This is likely due to variable levels of different adhesion molecules on acute or chronically activated HUVECs. These preliminary results suggest that the Endothelium-on-a-chip, as partner in novel therapeutic studies, could help monitoring dynamics of targeted

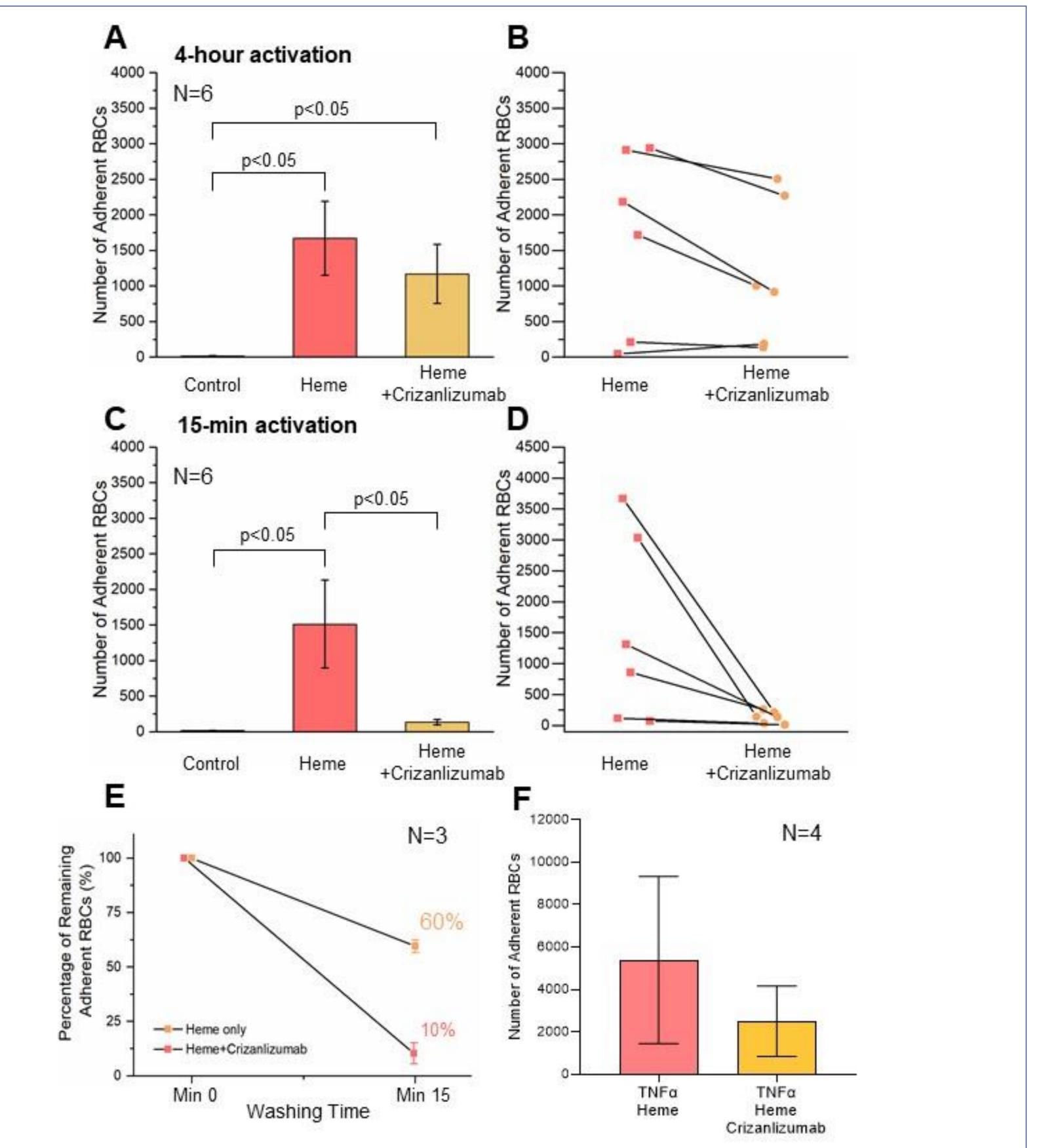


Figure 3. Effect of Crizanlizumab on SCD RBC adhesion to heme-activated HUVECs. (A) 4-hours heme activation statistically significant increases RBC adhesion to HUVECs while pre-incubation of heme-activated HUVECs via Crizanlizumab has a mild effect in lowering the RBC adhesion levels. (B) RBC adhesion to 4-hours heme-activated HUVECs has a strong subject-specific dependency. (C) Crizanlizumab treatment of 15-minutes heme-activated HUVECs resulted in statistically significant reduced RBC adhesion levels. (D) Inhibition of RBC adhesion was largely independent of subject-specific baseline adhesion levels. (E) RBCs were first allowed to adhere to 15-min heme-activated HUVECs and then rinsed with a solution containing either only heme or both heme and Crizanlizumab. The Crizanlizumab containing solution resulted in 90% of the adherent cells to detach after a 15-minute rinse while 60% of the RBCs remained adherent after a 15-minutes rinse with the solution that did not contain Crizanlizumab. (F) The short-term heme activation was preceded by 4-hours treatment with TNFα. Crizanlizumab reduce the number of adherent RBCs to the HUVECs pretreated with TNFa and heme. For all experiments, we used the following concentrations heme 40 μ M, Crizanlizumab 100 μ g/ml and TNF α 20 ng/ml. P-values were based on student's t-test. Error bars represent the standard error of the mean (SEM).

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